Periimplantitis and Implant Body Roughness: A Systematic Review of Literature

Fabienne Jordana, MSc, DDS, PhD,* Léa Susbielles, DDS,† and Jacques Colat-Parros, MSc, DDS, PhD,*

eriimplantitis is an infectious process that occurs after osseointegration of the implant, thus after the formation of a functional interface between the bone and the implant.¹ Periimplant inflammation affects the surrounding hard and soft tissue.² Periimplantitis develops progressively from inflammation around the implant, which will increase in the soft tissues, eventually causing marginal bone loss.³ Periimplantitis affects 16% to 28% of patients implanted, in the short or long term.^{4,5} Bacteriologically, many germs are found in the infected periimplant site but they are mainly anaerobes (Aggregatibacter actinomycetemcomitans, Porphyromonas gingivalis, Prevotella intermedia, Tannerella forsythia. Treponema denticola, etc).^{6–10} Some authors think that this disease entity is a foreign body reaction rather than an infective process.^{11,12} The multiple causes of periimplantitis warrant better analysis.

An important parameter for the clinical success of dental implants is the formation of a direct bone-implant

*Associate Professor, Dental Faculty, University of Nantes, Nantes, France, Hospital Practitioner, Dentistry Department, University Health Centre, Nantes, France. †Private Practice, Pau, France. ±Hospital Practitioner, Dentistry Department, Pellegrin Hospital, University Health Centre, Bordeaux, France; Associate

Reprint requests and correspondence to: Fabienne Jordana, MSc, DDS, PhD, University of Nantes, 1 Place Alexis Ricordeau, BP 84215, 44042 Nantes cedex 1, France, Phone: +33670580024, Fax: +332 40 20 18 67, E-mail: fabienne.jordana@univ-nantes.fr

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Purpose: The aim of this systematic review was to evaluate whether implant roughness is associated with periimplantitis in humans.

Materials and Methods: An electronic search of 3 databases (MEDLINE, Web of Knowledge, and the Cochrane Library) was undertaken until October 2017 and was supplemented by manual searching. Prospective studies were included if they met the following criteria: (1) give a clear definition of periimplantitis and (2) contain outcome data (clinical and radiological data) considering the periimplantitis rate. A systematic review was carried out to evaluate the impact of roughness on the periimplantitis rate.

Results: Of 4690 potentially eligible articles, 22 were included in the qualitative analysis and quantitative synthesis.

Conclusions: This systematic review suggests that roughness and surface treatment of dental implants are important factors associated with periimplantitis. (Implant Dent 2018;27:672-681)

Key Words: periimplant disease, surface roughness, implant surface

contact (BIC) that is directly influenced by the implant's surface roughness.¹³ Implant surface topography at the micrometer level of resolution has been regarded as the most important factor for successful implant treatment.¹⁴ Surface topography influences wound healing after implantation and also affects osseointegration.^{15,16} Surface roughness measurement is the measurement of the small-scale variations in the height of a physical surface.¹⁷ Some parameters, such as arithmetic mean surface roughness (Sa), that is, the average height of the analyzed area (micrometer), were used to describe the surface topography. Sa represents the arithmetic mean of the roughness area from the mean plane, for the height of the peaks and valleys according to the ISO 25178 standard.¹⁸ Sa expresses, as an absolute value, the difference in height of each point compared to the arithmetical mean of the surface. According to Albrektsson and Wennerberg,¹⁹ implants may be classified into 4 types according to surface roughness: smooth (Sa $< 0.5 \,\mu$ m); minimally rough (Sa between 0.5 and $1.0 \,\mu$ m), moderately rough (Sa between 1.0 and 2.0 µm), and rough (Sa $> 2.0 \,\mu$ m).

The aim of this review was to examine whether implant roughness is associated with periimplantitis in humans through a literature review.

MATERIALS AND METHODS

The Preferred Reporting Items for Systematic Reviews and Meta-Analyses system^{20,21} was adopted for this systematic review.

Study Protocol and Criteria

The protocol was designed to answer the following question: "In

Professor, Dental Faculty, University of Bordeaux, Bordeaux, France.

subjects with dental implants, do the implant surface treatment or surface roughness characteristics lead to periimplantitis?" It included studies reporting on at least 15 participants, randomized clinical trials, prospective cohort studies, retrospective studies, case-control studies, and cross-sectional studies in humans after implantation. Periimplantitis was defined by Heitz-Mayfield²² as the radiographic presence of bone loss ≥ 2 mm since the time of prosthetic replacement, positive bleeding on probing, and probing depth ≥ 4 mm.

Inclusion criteria. Only studies published in English in an international peer-reviewed journal were included. The studies had to describe surface treatment or surface roughness characteristics that can lead to periimplantitis. They also had to give a clear definition of periimplantitis or contain clinical and radiological data, which the reviewers could reliably relate to periimplantitis. Exclusion criteria. In vitro studies and animal studies were excluded. The following PECO (Population, Exposure to risk factor, Comparison, Outcome) definitions were considered for systemic search:

- Population: studies had to include systemically healthy patients with implant treatment;
- Exposure: periimplant disease diagnosed under a clinical and radiographic examination and adhered to a previously described definition;
- Comparison: the specific comparisons investigated were differences either in implant surface characteristics or in implant roughness;
- Outcome measures: the primary outcome variable was periimplantitis.

Search Strategy

A literature search was performed in MEDLINE via the PubMed database of the US National Library of Medicine, in the Web of Science, and Cochrane Library databases as well as a hand search of other literature to identify articles of potential relevance. The



Fig. 1. Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) flow diagram demonstrating the results of the systematic literature search.



Fig. 2. Electronic databases used and journals searched manually for the systematic literature search.

search included articles accepted for publication from 2000 up to October 2017. Previously published review articles on similar topics were also analyzed to assess potentially relevant publications.

The following keywords were used for this purpose:

Table 1. Design of Selected Publications²⁴⁻⁴⁵

	Studied Implants						
Authors	Follow-up Period (y)	Patient Number	Manufacturer	Surface Treatment	Roughness	Imp Nun a Inclu	olant nber at usion
Behneke et al ²⁴	5	55	ITI TPS (Straumann)	TPS	Rough	114	
Eliasson et al ²⁵	5	119	Brånemark Systeme (Nobel Biocare)	Machined	Minimally	476	
Astrand et al ²⁶	3	28	ITI TPS (Straumann)	TPS	Rough	77	150
			Brånemark Systeme (Nobel Biocare)	Machined	Minimally rough	73	
Astrand et al ²⁷	5	68	TiOblast (Astra Tech)	Sandblasted	Moderately rough	184	371
			Brånemark Systeme (Nobel Biocare)	Machined	Minimally rough	187	
Karoussis et al ²⁸	10	89	ITI TPS (Straumann)	TPS	Rough	179	
Vroom et al ²⁹	12	20	TiOblast (Astra Tech)	Sandblasted	Moderately rough	40	80
			Dental system (Astra Tech)	Machined	Minimally rough	40	
Jacobs et al ³⁰	16	18	TiOBlast (Astra Tech)	Sandblasted	Moderately rough	50	95
D 1/21	10	000	Branemark System MK II (Nobel Biocare)	Machined	rough	45	
Buser et al ³	10	303	(Straumann Dental Implant System (Straumann)	acid-etched	Noderately rough	511	
Charyeva et al ³²	6	108	MIS System (MIS)	acid-etched	rough	324	
Eiseber and	10	04	CLA depted implant avetem		rough	110	
Stenberg ³⁴	10	24	(Straumann)	acid-etched	rough	20	
	10	20			rough	20	
Mertens et al ³⁶	11	17	TiOBlast (Astra Tech)	Sandblasted	Moderately rough	94	
Ostman et al ³⁷	11	46	TiUnite (Nobel Biocare)	Anodic oxidation	Moderately rough	121	
Schliephake et al ³⁸	5	44	Osseospeed (Astra Tech)	Sandblasted + acid-etched	Moderately rough	143	
Chappuis et al ³⁹	20	98	Bonefit (Straumann)	TPS	Rough	145	500
Polizzi et al40	6-10	122	Til Inita (Nobel Riccare)	Machined	rough	257	500
D 11 1 141	10.15	22			rough	243	074
Ravald et al41	12–15	66	TiOblast (Astra Tech)	Sandblasted	Moderately rough	184	371
			Brånemark System (Nobel Biocare)	Machined	Minimally rough	187	
Jungner et al ⁴²	5	103	TiUnite (Nobel Biocare)	Anodic oxidation	Moderately rough	154	287
			Brånemark MKIII (Nobel Biocare)	Machined	Minimally rough	133	
van Velzen et al ⁴³	10	250	Soft-Tissue Level SLActive (Straumann)	Sandblasted + acid-etched	Moderately rough	506	
Becker et al44	12–23	92	ITI TPS (Straumann)	TPS	Rough	388	
Glauser ⁴⁵	11	38	TiUnite MK IV (Nobel Biocare)	Anodic oxidation	Moderately rough	102	

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Table 2. Implant Systems Used in the Literature: Mo	rphology and T	opography ^{24–45}				
						Surface
Implants	Design	Connection	Titanium	Sa (µm)	Roughness	Treatment
Dental Implant System Astra Tech (Astra Tech)*,29	Cylindrical	Internal	Grade 4	0.5 < Sa < 1	Minimally rough	Machined
Osseospeed (Astra Tech)*,38	Conical or	Internal	Grade 4	1.5	Moderately rough	Sandblasted + acid-etched
	cylindrical					
Astra TiO ₂ -Blast (Astra Tech)*, ^{27,29,30,35,36,41}	Cylindrical	Internal	Grade 3	1.1	Moderately rough	Sandblasted
MIS Seven dental implant system (MIS implant	Conical	Internal	TA ₆ V ₄	≈1.7	Moderately rough	Sandblasted + acid-etched
technologies)† ^{,32}						
Brånemark Standard (Nobel Biocare)‡ ^{,40}	Cylindrical	External	Grade 1	0.5	Minimally rough	Machined
Brånemark Markll (Nobel Biocare) ^{‡,27,30,40,41}	Cylindrical	External	Grade 1	0.7	Minimally rough	Machined
Brånemark MarkIII (Nobel Biocare) ^{‡,40,42}	Cylindrical	External	Grade 4	0.9	Minimally rough	Machined
Brånemark MarkIV (Nobel Biocare)‡,40	Cylindrical	External	Grade 4	0.9	Minimally rough	Machined
Brånemark MarkIII TiUnite (Nobel	Cylindrical	External	Grade 4	1.1	Moderately rough	Anodic oxidation
Biocare)‡, ^{33,37,40,42,45}						
Brånemark MarkIV TiUnite (Nobel Biocare)‡,37,40,45	Cylindrical	External	Grade 4	1.1	Moderately rough	Anodic oxidation
ITI TPS (Straumann)§, ^{24,26,28,44}	Cylindrical	Internal	Grade 4	>2	Rough	TPS
Bonefit TPS (Straumann)§,39	Cylindrical	Internal	I	>2	Rough	TPS
ITI SLActive (Straumann)§, ^{31,34}	Cylindrical	Internal	Grade 4	1.75	Moderately rough	Sandblasted + acid-etched
Soft Tissue-Level SLActive (Straumann)§ ^{,43}	Cylindrical	Internal	Grade 4	1.75	Moderately rough	Sandblasted + acid-etched
*Astra Tech AB, Mölndal, Sweden.						

tMIS Implant Technologies Ltd, Israël. tNobel Biocare AB, Göreborg, Sweden. §Straumarn AG, Waldenburg, Switzerland Brånemark System (Nobel Biocare)‡ machined (MK not identified).^{25,28}

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- periimplantitis OR peri-implantitis OR peri implantitis OR periimplant OR periimplant diseases OR peri-implant disease OR peri implant disease
- AND titanium OR dental implant OR implant

AND

- 1. surface characteristic OR surface roughness OR material characteristic OR titanium surface OR implant type OR implant surface OR surface decontamination OR surface topography.
- surface treatment OR TPS OR titanium plasma-sprayed OR anodic oxidation OR SLA OR machined surface OR turned surface OR sandblasted and acid-etched surface OR sandblasted surface OR acid-etched surface.

Quality Assessment

Quality assurance was developed by independent screening by 2 reviewers (L.S., F.J.) according to Khan et al.²³ When disagreement arose in the selection and eligibility, it was resolved by discussion between the 2 reviewers.

Data Extraction and Synthesis

The search gave 4690 results. Two independent reviewers (L.S., F.J.) analyzed titles and abstracts during the first stage of screening. Irrelevant articles were discarded. Additional manual searching of reference lists in the articles selected and in a number of review articles was performed to source further relevant publications (Fig. 1). The implantology and periodontology journals were searched manually between 2000 and 2017: Clinical Implants Dentistry and Related Research, Clinical Oral Implants Research, European Journal of Oral Implantology, Journal of Oral Implantology, Implant Dentistry, International Journal of Oral and Maxillofacial Implants, Journal of Periodontology, Journal of Clinical Periodontology, and Periodontology 2000 (Fig. 2). Ninety-eight full-text articles were assessed for eligibility. The full texts of the articles were read to determine whether the studies fulfilled the predetermined inclusion criteria. Twenty-two studies fulfilled the inclusion criteria and 76 were excluded (Fig. 1).

Table 3. Periimpla	able 3. Periimplantitis Rates (%) Depending on the Roughness ²⁴⁻⁴⁵							
Implant Roughness	Authors	Periimplantitis Rate (%)	Periimplantitis Rate Depending on the Roughness (%)					
Minimally rough	Eliasson et al ²⁵	0.00	0.57					
, ,	Astrand et al ²⁶	0.00						
	Astrand et al ²⁷	0.00						
	Vroom et al ²⁹	0.00						
	Jacobs et al ³⁰	0.00						
	Polizzi et al40	0.39						
	Ravald et al41	3.20						
	Jungner et al ⁴²	0.65						
Moderately rough	Astrand et al27	1.09	3.43					
	Vroom et al ²⁹	0.00						
	Jacobs et al ³⁰	0.00						
	Buser et al ³¹	1.76						
	Charyeva et al ³²	3.70						
	Degidi et al ³³	6.20						
	Fischer and Stenberg ³⁴	3.92						
	Gotfredsen ³⁵	5.00						
	Mertens et al ³⁶	2.13						
	Östman et al ³⁷	1.65						
	Schliephake et al ³⁸	6.29						
	Polizzi et al40	3.70						
	Ravald et al41	4.30						
	Jungner et al ⁴²	2.26						
	van Velzen et al ⁴³	4.20						
	Glauser ⁴⁵	7.56						
Rough	Behneke et al ²⁴	12.28	12.86					
	Astrand et al ²⁶	11.69						
	Karoussis et al ²⁸	15.40						
	Chappuis et al ³⁹	20.00						
	Becker et al44	9.76						

Data were collated into tables (Tables 1–5) and grouped according to implant roughness and surface treatment. Data synthesis was performed based on the evidence tables alone, and the data were further interpreted. Statistical analyses (Chi2) were carried out using XLStat (Addinsoft).

RESULTS

The initial search of the literature up to October 2017 yielded 4690 potentially suitable articles. After the exclusion of reviews, animal and *in vitro* studies, and studies that inappropriately identified periimplantitis or surface treatment or roughness characteristics, 22 publications remained fully eligible for this review. A meta-analysis could not be performed because of the heterogeneity of the reviewed studies. The κ value for interviewer agreement for study inclusion was 0.92 for titles and abstracts and 1.00 for full-text articles, indicating strong agreement.

Periimplantitis Rate and Implant Survival Rate According to Roughness

For the minimally rough surfaces, the Sa is between 0.5 and 1 μ m. The mean periimplantitis rate observed is 0.57% (Table 3). Of 8 studies referenced,^{25–27,29,30,40–42} five^{25–27,29,30} do not present periimplantitis. Two studies^{40,42} present a low rate between 0.39% and 0.65%. Finally, the study by Ravald et al⁴¹ indicates a maximum rate of 3.20% for 184 implants.

For the moderately rough surfaces, the Sa is between 1 and 2 μ m. Our literature review covered 16 studies; periimplantitis rates are between 0.00% and 7.56%, with a mean rate of 3.43%. There is a high dispersion of results. For the rough surfaces, the Sa is greater than 2 μ m. The mean periimplantitis rate is 12.86%, with rates ranging between 9.76% and 20.00% according to the studies.

Therefore, the minimally rough surfaces have very low periimplantitis rates. The periimplantitis rate increases with the moderately rough surfaces according to their roughness and to the technique used to obtain the roughness. The rough surfaces have the highest periimplantitis rates, which can reach 20.00%.

The statistical analysis is statistically significant ($P < 10^{-6}$) and allows us to say that the higher the roughness is, the higher is the periimplantitis rate.

Periimplantitis Rate According to Surface Treatment

The minimally rough surfaces are obtained by machining. The moderately rough surfaces are obtained by sandblasting, sandblasting + acid etching, or by anodic oxidation (Table 4). The rough surfaces are obtained by titanium plasma-sprayed (TPS). There is a statically significant difference in the frequency of periimplantitis between the different surface treatments ($P = 10^{-6}$).

In our literature review, all the minimally rough surfaces are machined. The periimplantitis rates are between 0.00% and 3.20% with the machined surfaces.^{25–27,29,30,40–42} The mean periimplantitis rate with the machined surfaces is 0.57%.

For the moderately rough implant surfaces, a relative heterogeneity of the periimplantitis rates can be noted. The lowest rates are observed with the sandblasted surfaces with an Sa of 1.1 μ m. The mean periimplantitis rate with the sandblasted surfaces is 2.38% with rates ranging between 0.00% and 5.00%. These surfaces seem clinically favorable. When the surface is obtained by anodic oxidation, however, the Sa is also 1.1 μ m. The mean periimplantitis rate is 4.14%, with rates ranging between 1.65% and 7.56%.^{33,37,40,42,45}

The surfaces obtained by sandblasting + acid etching (SLA) have an Sa of 1.75 μ m. The mean periimplantitis rate is 3.41%, with rates ranging from 1.76% to 6.29%.

Table 4: Periimplantitis Rates (%) According to Surface Treatment ²⁴⁻⁴⁵						
Surface Treatment	Authors	Sa (µm)	Periimplantitis Rates (%)	Periimplantitis Rates According to Surface Treatment (%)		
Machined	Eliasson et al ²⁵	0.9	0.00	0.57		
	Astrand et al ²⁶	0.9	0.00			
	Astrand et al ²⁷	0.9	0.00			
	Vroom et al ²⁹	0.9	0.00			
	Jacobs et al ³⁰	0.7	0.00			
	Polizzi et al40	0.5-0.9	0.39			
	Ravald et al ³¹	0.7	3.20			
	Jungner et al ⁴²	0.9	0.65			
Sandblasted	Astrand et al ²⁷	1.1	1.09	2.38		
	Vroom et al ²⁹	1.1	0.00			
	Jacobs et al ³⁰	1.1	0.00			
	Gotfredsen ³⁵	1.1	5.00			
	Mertens et al ³⁶	1.1	2.13			
	Ravald et al41	1.1	4.30			
Anodic	Degidi et al ³³	1.1	6.20	4.14		
oxidation	Östman et al ³⁷	1.1	1.65			
	Polizzi et al ⁴⁰	1.1	3.70			
	Jungner et al42	1.1	2.26			
	Glauser ⁴⁵	1.1	7.56			
Sandblasted +	Buser et al ³¹	1.75	1.76	3.41		
acid-etched	Charyeva et al ³²	1.75	3.70			
	Fischer and Stenberg ³⁴	1.75	3.92			
	Schliephake et al ³⁸	1.75	6.29			
	van Velzen et al43	1.75	4.20			
TPS	Behneke et al ²⁴	>2	12.28	12.86		
	Astrand et al ²⁶	>2	11.69			
	Karoussis et al ²⁸	>2	15.40			
	Chappuis et al ³⁹	>2	20.00			
	Becker et al44	>2	9.76			

The highest periimplantitis rates are found with the surfaces treated by TPS. Their Sa is higher than 2 μ m. The mean periimplantitis rate is 12.86%, with rates ranging from 9.76% to 20.00%.

These studies confirm that the periimplantitis rate increases with surface roughness. The results differ, however, for the moderately rough surfaces. The sandblasted surfaces have an Sa close to that of the minimally rough surfaces. The surfaces treated by sandblasting + acid etching have higher periimplantitis rates than the sandblasted-only surfaces. Their Sa is 1.75 μ m. It is close to that of the rough surfaces. However, the surfaces treated by anodic oxidation, with an Sa of 1.1 μ m, have high periimplantitis rates, with a mean rate of 4.14%.

Periimplantitis Rate According to Follow-up

Six studies were carried out over periods ranging from 1 to 5 years (Table 5).^{24–27,38,42} All the studies observe a periimplantitis rate of 0.11% with theminimally rough surfaces over periods of 1 to 5 years, from 1.09% to 6.29% for moderately rough implants and from 11.69% to 12.28% for rough implants.

Eight studies^{28,31–35,40,43} present a follow-up of between 6 and 10 years. The periimplantitis rates are higher than for the studies carried out over a 5-year period. The periimplantitis rates observed with machined surfaces are 0.39% and with rough surfaces treated by TPS are 15.40%.²⁸ The periimplantitis rates range from 1.76% to 6.20% with the moderately rough surfaces.

studies^{29,30,36,37,39,41,44,45} Eight presenting a follow-up of more than 11 years were included in this literature review. The longest is that of Chappuis et al,³⁹ with a 20-year follow-up of TPS rough surfaces. This study indicates a 20.00% periimplantitis rate.³⁹ The minimally rough surfaces have periimplantitis rates that vary from 0.00% to 3.20%. The moderately rough surfaces give results ranging from 0.00% to 7.56%, with mean rate of 3.21%. The mean periimplantitis rate is less than that of the 5- to 10-year studies. The studies carried out over more than 11 years concern implants with surfaces treated by sandblasting or by anodic oxidation. No significant difference was found with respect to the moderately rough surfaces according to follow-up time (P = 0.81). For the moderately rough surfaces, a periimplantitis rate of 3.04% is observed, and then stabilization is observed over time.

The periimplantitis rate with the rough surfaces increases with the study time. The rate of occurrence of periimplantitis is 12.04% between 1 and 5 years, 15.40% between 6 and 10 years. At 20 years, the periimplantitis rate is 20% in the study by Chappuis et al.³⁹ For the minimally rough surfaces, however, the results of the 1- to 5-year studies and the 6- to 10-year studies are comparable, but there is a statistically significant increase in the periimplantitis rate after 11 years ($P = 1.2 \times 10^{-4}$). However, the periimplantitis frequency remains low.

DISCUSSION

There is no consensual definition of periimplantitis or of its clinical parameters. The 6th European Consensus Conference (2008) gave the following definition: "the lesion of periimplant mucositis resides in the soft tissues, periimplantitis also affects the supporting bone."3 Definitions of periimplantitis, which may include different clinical and radiological thresholds, vary because of the various thresholds of bone loss and pocket depths used in the literature.46 Correct diagnosis of periimplant disease is essential to appropriately manage periimplant disease.³² We used this definition of

Table 5. Periimplantitis Rates (%) According to Follow-up Period and Roughness ²⁴⁻⁴⁵							
Follow-up		Minir	nally	Mode	erately	_	
Period (y)	Authors	Rou	ugh	Roi	ugh	Roi	ugh
0–5	Behneke et al ²⁴	—	0.11	—	3.04	12.28	12.04
	Eliasson et al ²⁵	0.00		—		—	
	Astrand et al ²⁶	0.00		—		11.69	
	Astrand et al ²⁷	0.00		1.09		—	
	Schliephake et al ³⁸	—		6.29		—	
	Jungner et al ⁴²	0.65		2.26		—	
6–10	Karoussis et al ²⁸	—	0.39	—	3.59	15.40	15.40
	Buser et al ³¹	—		1.76			
	Charyeva et al ³²	—		3.70		_	
	Degidi et al ³³	—		6.20			
	Fischer and	—		3.92		—	
	Stenberg ³⁴						
	Gotfredsen ³⁵	—		5.00		—	
	Polizzi et al40	0.39		3.70		—	
	van Velzen et al ⁴³			4.20		—	
11–20	Vroom et al ²⁹	0.00	2.40	0.00	3.21	—	12.06
	Jacobs et al ³⁰	0.00		0.00		—	
	Mertens et al ³⁶	—		2.13		—	
	Östman et al ³⁷	—		1.65		—	
	Chappuis et al ³⁹	—		—		20.00	
	Ravald et al41	3.20		4.30		—	
	Becker et al44	—		—		9.76	
	Glauser ⁴⁵	—		7.56		—	

periimplantitis: radiographic presence of bone loss ≥ 2 mm since the time of prosthetic replacement, positive bleeding on probing, and probing depth ≥ 4 mm.²² According to Albrektsson and Wennerberg,¹⁹ implants may be classified into 4 types according to surface roughness: smooth (Sa < 0.5 µm); minimally rough (Sa between 0.5 and 1.0 µm), moderately rough (Sa between 1.0 and 2.0 µm), and rough (Sa > 2.0 µm). Sa represents the arithmetic mean of the roughness area from the mean plane, for the height of the peaks and valleys.¹⁸

The first implants to be produced (machined or turned) are still considered the gold standard for implant surfaces. Their minimally rough surfaces (Sa typically ranging from 0.4 to 0.8 μ m) have periodic grooves. In the 2000s, rougher surfaces were sought to increase the BIC, improve implant stability, and allow earlier implant loading.^{47,48} Some authors considered that the increased surface roughness of commercially pure titanium implants would improve BIC and the mechanical properties of the interface^{49–53} and that the

improved platelet activation could positively regulate the osteogenic responses.⁵⁴

The majority of currently marketed implants are moderately rough (Sa between 1.0 and 2.0 µm).²² Albrektsson and Wennerberg¹⁹ showed that there is an optimum surface roughness window from 1 to 1.5 µm, for which there is a compromise between engineering and clinical practice. They consider that a higher value leads to a loss of bone anchoring.¹⁹ Quirynen et al⁵⁵ report that implants with relatively smooth surfaces must be used to prevent biological complications. Many studies^{56–58} show that rough implants develop significantly more periimplantitis. Esposito et al⁵⁹ showed that the use of machined implants (minimally rough surfaces) instead of rough implants could bring a 20% reduction in the periimplantitis rate. These authors confirm the results of our literature review.

Surface treatments influence implant roughness. Machined implants have a relatively smooth surface.⁶⁰ Sandblasting consists of forcing small grits of chosen shape and size across implant surfaces, usually by compressed air.⁶⁰ Acid etching by immersion in strong acids creates a microroughness with irregular pits of varying depths on the surface.⁶⁰ With the SLA method, the implant surface is first sandblasted with large grit, then the acid etching forms micropits on its surface. The rough implants are all produced using the TPS technique. TPS dental implants have a complex surface; the particle density in the valleys normally appears higher than those on the thread peaks.⁶¹ We have not included hydroxyapatite-coated implants in our literature review because there are many controversies about their longterm prognosis.

The sandblasted + acid-etched surfaces have an Sa of 1.75 μ m, and the rough surfaces have an Sa greater than 2 μ m. In our literature review, the mean periimplantitis rate with surfaces treated by sandblasting + acid etching is 3.41%. The bone loss observed with the machined or sandblasted implants is equivalent, as with the occurrence of periimplantitis according to several authors.^{62–64} BIC is even greater with sandblasted surfaces than with machined surfaces.^{19,65–67}

In our systematic review of literature, 2 studies^{29,30} compare sandblasted surfaces and machined surfaces. No significant difference is observed. The periimplantitis rate is 0% in both these studies. The mean periimplantitis rate with surfaces treated by anodic oxidation (moderately rough surfaces) is 4.14%. The study by Polizzi et al⁴⁰ concludes that these surfaces have a higher periimplantitis rate (3.7%) than the machined surfaces (0.39%). The surfaces treated by anodic oxidation have an Sa equivalent to the sandblasted surfaces $(1.1 \,\mu\text{m})$; their periimplantitis rate is, however, higher. The difference is due to their surface treatment. The sandblasted surface is obtained by subtraction, and the surface treated by anodic oxidation is obtained by addition. Finally, implants with a very rough TPS surface have periimplantitis rates that can reach 20.00%. It should be noted that the periimplantitis rates observed with TPS surfaces (12.86%) are, in our literature review, lesser to those of the study by Dam et al^{68} (18.00%).

Periimplantitis is one of the major problems in implantology. Many uncertainties remain regarding its etiopathogenesis. Bacterial infection is an aggravating factor. Also, the role of the operator and the host's response cannot be excluded. Our literature review shows that surface roughness plays a major role in the occurrence of periimplantitis.

CONCLUSION

Periimplantitis is clearly linked with surface roughness according to the results of our systematic review of literature. The higher the surface roughness, the higher the mean periimplantitis rate. Up to an arithmetic mean surface roughness (Sa) of 1 μ m, there is little periimplantitis. Periimplantitis appears for Sa values greater than 1.2 μ m.

Considering the reviewed studies as a whole, it is evident that implant roughness is associated with periimplantitis. Although a comparison of the published results was limited due to the lack of homogeneity of the studies, it is clear that clinicians should give priority to the use of implants with machined or even sandblasted surfaces.

DISCLOSURE

The authors claim to have no financial interest, either directly or indirectly, in the products or information listed in the article.

ROLES/CONTRIBUTIONS BY AUTHORS

F. Jordana made substantial contributions to conception and design, acquisition of data, or analysis and interpretation of data; drafted or critically revised the manuscript for important intellectual content; and provided final approval of the manuscript. L. Susbielles made substantial contributions to conception and design, acquisition of data, or analysis and interpretation of data; drafted or critically revised the manuscript for important intellectual content; and provided final approval of the manuscript. J. Colat-Parros made substantial contributions to conception and design, acquisition of data, or analysis and interpretation of data; drafted or critically revised the manuscript for important intellectual content; and provided final approval of the manuscript.

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F. Jordana and L. Susbielles contributed equally to this work.

REFERENCES

1. Mouhyi J, Dohan Ehrenfest DM, Albrektsson T. The peri-implantitis: Implant surfaces, microstructure, and physicochemical aspects. *Clin Implant Dent Relat Res.* 2012;14:170–183.

2. Smeets R, Henningsen A, Jung O, et al. Definition, etiology, prevention and treatment of peri-implantitis: A review. *Head Face Med.* 2014;10:34.

3. Lindhe J, Meyle J; Group D of European Workshop on Periodontology. Peri-implant diseases: Consensus report of the sixth European Workshop on Periodontology. *J Clin Periodont.* 2008; 35:282–285.

4. Fransson C, Lekholm U, Jemt T, et al. Prevalence of subjects with progressive bone loss at implants. *Clin Oral Implants Res.* 2005;16:440–446.

5. Rinke S, Ohl S, Ziebolz D, et al. Prevalence of peri-implant disease in partially edentulous patients: A practice based cross-sectional study. *Clin Oral Implants Res.* 2011;22:826–833.

6. Pontoriero R, Tonelli MP, Carnevale G, et al. Experimentally induced peri-implant mucositis: A clinical study in humans. *Clin Oral Implants Res.* 1994;5:254–259.

7. Mombelli A, Lang NP. The diagnosis and treatment of peri-implantitis. *Periodontology 2000.* 1998;17:63–76.

8. Leonhardt A, Renvert S, Dahlén G. Microbial findings at failing implants. *Clin Oral Implants Res.* 1999;10:339–345.

9. Teughels W, Van Assche N, Sliepen I, et al. Effect of material characteristics and/or surface topography on biofilm. *Clin Oral Implants Res.* 2006;17:68–81.

10. Hultin M, Gustafsson A, Hallström H. Microbiological findings and host response in patient with peri-implantitis. *Clin Oral Implants Res.* 2012;13:349–358.

11. Albrektsson T, Dahlin C, Jemt T, et al. Is marginal bone loss around oral implants the result of a provoked foreign body reaction? *Clin Implant Dent Relat Res.* 2014;16:155–165.

12. Albrektsson T, Canullo L, Cochran D, et al. Peri-implantitis: A complication of a foreign body or a man-made "disease." Facts and fiction. *Clin Implant Dent Relat Res.* 2016;18:840–849.

13. Rosa MB, Albrektsson T, Francischone CE, et al. The influence of surface treatment on the implant roughness pattern. *J Appl Oral Sci.* 2012;20:550–555.

14. Wennerberg A, Albrektsson T. Implant surfaces beyond micron roughness. Experimental and clinical knowledge of surface topography and surface chemistry. *Appl Osseointegration Res.* 2006;5:40–44.

15. Albrektsson T, Brånemark PI, Hansson HA, et al. Osseointegrated titanium implants: Requirements for ensuring a long-lasting, direct bone-toimplant anchorage in man. *Acta Orthop.* 1981;52:155–170.

16. Bauer S, Schmuki P, von der Mark K, et al. Engineering biocompatible implant surfaces. Part I: Materials and surfaces. *Prog Mater Sci.* 2013;58:261–326.

17. Butler DL. Surface roughness measurement. In: Li D, ed. *Encyclopedia of Microfluidics and Nanofluidics*. Boston, MA: Springer; 2014.

18. International Organization for Standardization. ISO 25178: Geometric Product Specifications (GPS)—Surface Texture: Areal—Part 2: Terms, Definitions and Surface Texture Parameters. Geneva, Switzerland; 2012:1–47.

19. Albrektsson T, Wennerberg A. Oral implant surfaces: Part 1: Review focusing on topographic and chemical properties of different surfaces and in vivo responses to them. *Int J Prosthodont*. 2004;17:536–543.

20. Transparent reporting of systematic reviews and meta-analyses. Available at: http://www.prisma-statement.org. Accessed October 21, 2017.

21. Moher D, Liberati A, Tetzlaff J, et al. Preferred reporting items for systematic reviews and meta-analyses: The PRISMA statement. *PLoS Med.* 2009;6:e1000097.

22. Heitz-Mayfield L. Peri-implant diseases: Diagnosis and risk indicators. *J Clin Periodont.* 2008;35:292–304.

23. Khan KS, ter Riet G, Popay J, et al. Stage II: Conducting the review. Phase 5: Study quality assessment. In: Khan KS, ter Tiet G, Glanvill J, et al, ed. *Undertaking Systematic Reviews of Research on Effectiveness.* 2nd ed. York, United Kingdom: University of York; 2001:1–20.

24. Behneke A, Behneke N, d'Hoedt B. The longitudinal clinical effectiveness of ITI solid-screw implants in partially edentulous patients: A 5-year follow-up report. *Int J Oral Maxillofac Implants.* 2000;15: 633–645.

25. Eliasson A, Palmqvist S, Svenson B, et al. Five-year results with fixed complete-arch mandibular prostheses supported by 4 implants. *Int J Oral Max-illofac Implants.* 2000;15:505–510.

26. Astrand P, Engquist B, Anzén B, et al. A three-year follow-up report of

a comparative study of ITI Dental Implants and Brånemark System implants in the treatment of the partially edentulous maxilla. *Clin Implant Dent Relat Res.* 2004;6: 130–141.

27. Astrand P, Engquist B, Dahlgren S, et al. Astra Tech and Brånemark system implants: A 5-year prospective study of marginal bone reactions. *Clin Oral Implants Res.* 2004;15:413–420.

28. Karoussis IK, Brägger U, Salvi GE, et al. Effect of implant design on survival and success rates of titanium oral implants: A 10-year prospective cohort study of the ITI dental implant system. *Clin Oral Implants Res.* 2004;15:8–17.

29. Vroom MG, Sipos P, de Lange GL, et al. Effect of surface topography of screw-shaped titanium implants in humans on clinical and radiographic parameters: A 12-year prospective study. *Clin Oral Implants Res.* 2009;20:1231–1239.

30. Jacobs R, Pittayapat P, van Steenberghe D, et al. A split-mouth comparative study up to 16 years of two screw-shaped titanium implant systems. *J Clin Periodont.* 2010;37:1119–1127.

31. Buser D, Janner SF, Wittneben JG, et al. 10-year survival and success rates of 511 titanium implants with a sandblasted and acid-etched surface: A retrospective study in 303 partially edentulous patients. *Clin Implant Dent Relat Res.* 2012;14:839–851.

32. Charyeva O, Altynbekov K, Zhartybaev R, et al. Long-term dental implant success and survival—A clinical study after an observation period up to 6 years. *Swed Dent J.* 2012;36:1–6.

33. Degidi M, Nardi D, Piattelli A. 10year follow-up of immediately loaded implants with TiUnite porous anodized surface. *Clin Implant Dent Relat Res.* 2012; 14:828–838.

34. Fischer K, Stenberg T. Prospective 10-year cohort study based on a randomized controlled trial (RCT) on implantsupported full-arch maxillary prostheses. Part 1: Sandblasted and acid-etched implants and mucosal tissue. *Clin Implant Dent Relat Res.* 2012;14:808–815.

35. Gotfredsen K. A 10-year prospective study of single tooth implants placed in the anterior maxilla. *Clin Implant Dent Relat Res.* 2012;14:80–87.

36. Mertens C, Steveling HG, Stucke K, et al. Fixed implant-retained rehabilitation of the edentulous maxilla: 11-year results of a prospective study. *Clin Implant Dent Relat Res.* 2012;14:816–827.

37. Östman PO, Hellman M, Sennerby L. Ten years later. Results from a prospective single-center clinical study on 121 oxidized (TiUnite) Brånemark implants in 46 patients. *Clin Implant Dent Relat Res.* 2012;14:852– 860. 38. Schliephake H, Rödiger M, Phillips K, et al. Early loading of surface modified implants in the posterior mandible—5-year results of an open prospective non-controlled study. *J Clin Periodont.* 2012; 39:188–195.

39. Chappuis V, Buser R, Brägger U, et al. Long-term outcomes of dental implants with a titanium plasma-sprayed surface: A 20-year prospective case series study in partially edentulous patients. *Clin Implant Dent Relat Res.* 2013;15:780–790.

40. Polizzi G, Gualini F, Friberg B. A two-center retrospective analysis of long-term clinical and radiologic data of TiUnite and turned implants placed in the same mouth. *Int J Prosthodont.* 2013;26:350–358.

41. Ravald N, Dahlgren S, Teiwik A, et al. Long-term evaluation of Astra Tech and Brånemark implants in patients treated with full-arch bridges. Results after 12 to 15 years. *Clin Oral Implants Res.* 2013;24:1144–1151.

42. Jungner M, Lundqvist P, Lundgren S. A retrospective comparison of oxidized and turned implants with respect to implant survival, marginal bone level and peri-implant soft tissue conditions after at least 5 years in function. *Clin Implant Dent Relat Res.* 2014;16:230–237.

43. van Velzen FJ, Ofec R, Schulten EA, et al. 10-year survival rate and the incidence of peri-implant disease of 374 titanium dental implants with a SLA surface: A prospective cohort study in 177 fully and partially edentulous patients. *Clin Oral Implants Res.* 2015;26:1121–1128.

44. Becker ST, Beck-Broichsitter BE, Rossmann CM, et al. Long-term survival of straumann dental implants with TPS surfaces: A retrospective study with a follow-up of 12 to 23 years. *Clin Implant Dent Relat Res.* 2016;18:480–488.

45. Glauser R. Implants with an oxidized surface placed predominately in soft bone quality and subjected to immediate occlusal loading: Results from an 11-year clinical follow-up. *Clin Implant Dent Relat Res.* 2016;18:429–438.

46. Charalampakis G, Jansåker E, Roos-Jansåke AM. Definition and prevalence of peri-implantitis. *Curr Oral Health Rep.* 2014;1:239–250.

47. Roccuzzo M, Bunino M, Prioglio F, et al. Early loading of sandblasted and acid-etched (SLA) implants: A prospective split-mouth comparative study. *Clin Oral Implants Res.* 2001;12:572–578.

48. Bornstein MM, Lussi A, Schmid B, et al. Early loading of nonsubmerged titanium implants with a sandblasted and acid-etched (SLA) surface: 3-year results of a prospective study in partially edentulous patients. Int J Oral Maxillofac Implants. 2003;18:659–666.

49. Cooper LF. A role for surface topography in creating and maintaining bone at titanium endosseous implants. *J Prosthet Dent.* 2000;84:522–534.

50. Class L, Wilke HJ, Steinemann S. The influence of various titanium surfaces on the interface shear strength between implants and bone. *J Biomech.* 1991;24: 461.

51. Wennerberg A, Ektessabi A, Albrektsson T, et al. A 1-year follow-up of implants of differing surface roughness placed in rabbit bone. *Int J Oral Maxillofac Implants*. 1997;12:486–494.

52. Buser D, Broggini N, Wieland M, et al. Enhanced bone apposition to a chemically modified SLA titanium surface. *J Dent Res.* 2004;83:529–533.

53. Ferguson SJ, Broggini N, Wieland M, et al. Biomechanical evaluation of the interfacial strength of a chemically modified sandblasted and acid-etched titanium surface. *J Biomed Mater Res A.* 2006;78: 291–297.

54. Park JY, Gemmell CH, Davies JE. Platelet interactions with titanium: Modulation of platelet activity by surface topography. *Biomaterials.* 2001;22: 2671–2682.

55. Quirynen M, De Soete M, Van Steenberghe D. Infectious risks for oral implants: A review of the literature. *Clin Oral Implants Res.* 2002;13:1–19.

56. Moberg LE, Köndell PA, Sagulin GB, et al. Brånemark system and ITI dental implant system for treatment of mandibular edentulism. A comparative randomized study: 3-year follow-up. *Clin Oral Implants Res.* 2001;12:450–461.

57. Marrone A, Lasserre J, Bercy P, et al. Prevalence and risk factors for periimplant disease in Belgian adults. *Clin Oral Implants Res.* 2013;24:934–940.

58. Quirynen M, Abarca M, Van Assche N, et al. Impact of supportive periodontal therapy and implant surface roughness on implant outcome in patients with a history of periodontitis. *J Clin Periodont.* 2007;34: 805–815.

59. Esposito M, Ardebili Y, Worthington HV. Interventions for replacing missing teeth: Different types of dental implants. *Cochrane Database Syst Rev.* 2014;22: CD003815.

60. Yan Guo C, Tin Hong Tang A, Pekka Matinlinna J. Insights into surface treatment methods of titanium dental implants. *J Adhes Sci Technol.* 2012;26: 189–205.

61. Klokkevold PR, Johnson P, Dagrostari S, et al. Early endosseous integration enhanced by dual acid etching of titanium: A torque removal study in the

rabbit. Clin Oral Implants Res. 2001;12: 350-357.

62. Wennström JL, Ekestubbe A, Gröndahl K, et al. Oral rehabilitation with implant-supported fixed partial dentures in periodontitis-susceptible subjects. A 5-year prospective study. *J Clin Periodont.* 2004;31:713–724.

63. Arnhart C, Dvorak G, Trefil C, et al. Impact of implant surface topography: A clinical study with a mean functional loading time of 85 months. *Clin Oral Implants Res.* 2013;24:1049–1054. 64. Welander M, Abrahamsson I, Berglundh T. Subcrestal placement of two-part implants. *Clin Oral Implants Res.* 2009;20:226–231.

65. Thomas KA, Cook SD. An evaluation of variables influencing implant fixation by direct bone apposition. *J Biomed Mater Res.* 1985;19:875–901.

66. Buser D, Schenk RK, Steinemann SG, et al. Influence of surface characteristics on bone integrations of titanium implants: A histomorphometric study in miniature pigs. *J Biomed Mater Res.* 1991;25:889–902.

67. Ericsson I, Johansson CB, Bystedt H, et al. A histomorphometric evaluation of bone-to-implant contact on machineprepared and roughened titanium dental implants. A pilot study in the dog. *Clin Oral Implants Res.* 1994;5:202–206.

68. Dam HG, Najm SA, Nurdin N, et al. A 5- to 6-year radiological evaluation of titanium plasma sprayed/sandblasted and acid-etched implants: Results from private practice. *Clin Oral Implants Res.* 2013;25: e159–e165.